REVIEW ARTICLE

Interleukin-10 and Coronary Disease

Ruth Pérez Fernández and Juan Carlos Kaski

Coronary Artery Disease Research Unit. Cardiological Sciences. St. George's Hospital Medical School. Londob. United Kingdom.

Understanding of the pathophysiology of atherosclerosis has changed markedly over the past few decades. It is now widely accepted that inflammation plays a fundamental role in the genesis and development of atherosclerosis. Inflammatory mechanisms also appear to determine clinical presentation and disease outcome. Atherosclerotic lesions have high concentrations of inflammatory cells (T lymphocytes and activated macrophages) as well as an abundance of pro-inflammatory cytokines [interleukin (IL)-1, IL-6, IL-8, interferon-γ, tumor necrosis factor-α, etc.] that modulate local inflammatory responses. These may also alter plaque stability and facilitate the development of acute cardiovascular events. The role of anti-inflammatory cytokines in this context remains to be studied. IL-10 is an anti-inflammatory cytokine synthesised by T-lymphocytes and macrophages and has other anti-inflammatory effects. IL-10 expression within human atherosclerotic plaques has been demonstrated and animal experiments have shown that low levels of IL-10 lead to the development of extensive and unstable atherosclerotic lesions. Currently available evidence suggests a potential protective role for IL-10 in atherosclerosis. This new perspective on coronary disease as a chronic inflammatory process may open new avenues for the management of ischemic heart disease.

Key words: Inflammation. Atherosclerosis. Interleukin-10.

Interleucina-10 y enfermedad coronaria

Los conocimientos actuales de la fisiopatología de la aterosclerosis divergen marcadamente de los de las últimas décadas. Hoy día se acepta de manera generalizada que la inflamación desempeña un papel fundamental en el desarrollo y progresión de las lesiones ateroscleróticas, condicionando la aparición de manifestaciones clínicas en la evolución. El estudio histopatológico de las lesiones ateroscleróticas revela la presencia de células inflamatorias (linfocitos T y macrófagos activados), así como de abundantes citocinas proinflamatorias (IL-1, IL-6, IL-8, TNF-α, INF-γ, etc.), que modulan la respuesta inflamatoria local, alterando la estabilidad de la placa y favoreciendo el desarrollo de acontecimientos cardiovasculares agudos. Sin embargo, el papel de las citocinas antiinflamatorias no ha sido tan bien estudiado. La IL-10 es una citocina antiinflamatoria capaz de inhibir la síntesis de citocinas proinflamatorias por los linfocitos T y los macrófagos, así como otras funciones inflamatorias de estas células. Su presencia ha sido demostrada en las placas ateroscleróticas humanas y se ha objetivado en estudios de experimentación animal que los bajos valores de IL-10 condicionan el desarrollo de lesiones ateroscleróticas más extensas y morfológicamente más inestables. Las evidencias disponibles en la actualidad sugieren un potencial papel protector de la IL-10 en el desarrollo de la aterosclerosis. Este nuevo enfoque de la enfermedad coronaria como una enfermedad inflamatoria crónica puede abrir en el futuro nuevos caminos para la investigación en el campo de la cardiopatía isquémica.

Palabras clave: Inflamación. Aterosclerosis. Interleucina-10.

INTRODUCTION

Coronary artery disease (CAD) is highly prevalent and one of the most important causes of morbidity and mortality in industrialized societies.^{1,2} The underlying process in CAD is atherosclerosis, and it is currently

Correspondence: Dr. Prof. J.C. Kaski. Head of Coronary Artery Disease Research Unit.

Cardiological Sciences, St. George's Hospital Medical School. Cranmer Terrace, London SW17 0RE.United Kingdom. E-mail: jkaski@sghms.ac.uk

considered a chronic inflammatory disease of the arterial wall.3-5 The most severe clinical presentation of this process is an acute coronary syndrome (unstable angina and infarcts $[AMI]$, $6-8$ which occurs secondary to the occlusion of the diseased arteries.

Histological study of the atherosclerotic plaques reveals the presence of progressive infiltration and accumulation of lipids, inflammatory cells (monocytes/macrophages, T-lymphocytes), smooth muscle cells (SMC), and an extracellular matrix in the arterial wall $1,2$

The identification of the inflammatory cells in the atherosclerotic lesions, as well as the complementary fac-

ABBREVIATIONS

DIC: disseminated intravascular coagulation. SMC: smooth muscle cell. CMV: cytomegalovirus. COX-2: cyclooxigenase-2. APC: antigen presenting cells. ED: endothelial dysfunction. DNA: deoxyribonucleic acid. CAD: coronary artery disease. TF: tissular factor. G-CSF: granulocyte colony stimulant factor. GM-CSF: granulocyte and monocyte colony stimulant factor. HSV: herpes virus simplex. AMI: acute myocardial infarction. ICAM-1: intracellular adhesion molecule-1. IL: interleukine. INF-γ: interferon γ. iNOS: nitric oxide synthetase. LDL: low density lipoproteins. MM-LDL: minimally modified LDL. ox-LDL: oxidized LDL. LPS: lipopolysacharide. MHC-II: major histocompatability complex class II molecules. MCP-1: monocyte chemoattractant protein-1. M-CSF: macrophage colony-stimulating factor. ECM: extracellular matrix. MMP: metalloproteinase. NF-κβ: nuclear factor κβ. NO: nitric oxide. RPC: reactive protein-C. PDGF: plaque-derived growth factor. PGI2: prostaglandin I2. mRNA: messenger ribonucleic acid. MPTI: metalloproteinase tissular inhibitor. Th1: type 1 T-helper cells. TNF-α: tumor necrosis factor-α. VCAM-1: vascular cell adhesion molecule. VEGF: vascular endothelial growth factor.

tors, immunoglobulins, cytokines, $3,9$ and others, implicates the involvement of the immunological system in atherogenesis. During this inflammatory reaction, a great quantity of cytokines is produced by macrophages and activated T-cells present in the plaque,³ charged with modulating inflammatory response. This process may alter plaque stability and favor the development of acute events.³ Nevertheless, the manner in which this local or systemic, or both, immunological response is initiated and propagated to produce or favor the development of atherosclerotic lesions is still not completely clear.

Numerous studies of experimental animals fed with diets rich in cholesterol have shown that immunosuppression causes the development of more extensive and more severe atherosclerotic lesions as compared to controls.¹⁰⁻¹²

In recent years, multiple scientific studies have emphasized the role of the immunological system cells (monocytes, lymphocytes, etc.) and pro-inflammatory cytokines (TNF-α, IL-1β, IL-6, INF-γ, etc.)^{3,13-17} in the development of atherosclerosis. Nevertheless, there is little evidence available on the potential role of antiinflammatory cytokines in this process.

The aim of this article is to review the knowledge currently available concerning the potential protective role of anti-inflammatory cytokines, specifically interleukine 10 (IL-10), in the pathogenesis and development of atherosclerotic lesions.

PHYSIOPATHOLOGY OF ATHEROSCLEROSIS

The development of atherosclerotic lesions is a process that begins in the second or third decades in the life of an individual, and can be divided into various stages through which the composition of the atherosclerotic plaque changes progressively until it acquires the morphology of a mature plaque.

Endothelial dysfunction

The first event in the development of atherosclerosis is the appearance of endothelial dysfunction (ED) .^{3,18-21} The endothelium plays an important part in maintaining the equilibrium of vascular bed function. It has a regulatory role in vasomotor tone by the production of vasodilator substances, such as nitric oxide (NO) and prostacycline (PGI2), and also of vasoconstrictor substances such as endothelin 1 and angiotensin II^{22} It also possesses anti-atherogenic (antiaggregate, anti-adhesive, anti-proliferative and antioxidant) and anti-inflammatory properties, segregating chemoattractant substances from monocytes and lymphocytes, as well as modulators of vascular growth.23-26

There are multiple causes of ED that favor the development of atherosclerosis including the presence of elevated modified LDL values (ox-LFL, MM-LDL); free radicals; immunoregulatory substances (TNF-α, IL-1β, LPS); infectious microorganisms (HSV, *Chlamydia*, CMV, etc.); genetic alterations; elevated serum homocysteine values, and classic risk factors (hypertension, diabetes, smoking).27,28

Endothelial dysfunction leads to a loss of the homeostatic functions of the endothelium, resulting in the adhesion of plaques and inflammatory cells (monocytes and lymphocytes to the vascular wall;²⁷ an increase in endothelial permeability that allows the deposit of modified LDL at the intimal level; 29 a liberation of cytokines and growth factors that produces the proliferation of smooth muscle cells and the attraction of more inflammatory cells to the altered arterial wall.²⁷ As a consequence, it also causes perturbation of the thrombolitic-thrombotic equilibrium in the endothelial bed that promotes the development of thrombotic phenomena, such as the abnormal regulation of vasomotor tone secondary to decreased bioavailability of NO, with the subsequent tendency to arterial vasoconstriction.^{30,31}

The role of low-density lipoproteins (LDL). The formation of fat striae

The first histopathological change detectable in the early phases of atherosclerosis is the accumulation of LDL particles in the subintimal space.³² These LDL undergo an oxidation process that activates the endothelium, favoring the development of atherosclerotic plaque.33,34 The modified (oxidated) LDL cause the expression of adhesion molecules $(ICAM-1, VCAM-1)³⁵$ and the synthesis of monocyte chemoattractant protein-1 (MCP-1) by the endothelial cells. This favors the union of the circulating monocytes and lymphocytes to the dysfunctional endothelium and later the migration of these cells to the sub endothelial space, promoting, at the same time, the differentiation of monocytes to macrophages.3,36 The oxidated LDL also change the production of free radicals and NO, favoring oxidative stress in the arterial wall,^{3,36,37} increasing the apoptosis of the endothelial cells.³⁸

The monocytes that are attracted to the dysfunctional endothelium by liberated chemoattractants adhere to the wall via the adhesion molecules (ICAM-1, VCAM-1) expressed by the damaged endothelial cells, internalize in the sub endothelial space and mature into macrophages, which capture the ox-LDL to transform themselves into frothy cells, initiating in this way the formation of fat striae.³

These cells loaded with lipids produce free radicals for the specific oxidation of LDL and free new cytokines for the attraction of more monocytes and lymphocytes to the dysfunctional endothelium and for the migration and proliferation of SMC in the intima.34,39-41 These processes auto perpetuate the mechanism that favors the development and progression of atherosclerotic plaque.

Vulnerability of atherosclerotic plaque

Atheromas are dynamic structures where equilibrium exists between the destructive influence of the inflammatory cells and the stabilizing effect of SMC.6,42 The latter are responsible for synthesizing extracellular matrix (ECM) proteins, the principal component of the fibrous cover of atherosclerotic plaques that gives the lesion stability.³⁶ In atherosclerotic plaques there is a balance between the processes of synthesis and collagen degradation that are narrowly controlled by the inflammation mediators and regulate the contents of same in atherosclerotic lesions.⁴³

The vulnerable plaque (that have a tendency to rupture) are characterized by a highly lipidic nucleus, an elevated infiltration of inflammatory cells (macrophages and T-lymphocytes), few SMC, and a thin fibrous cover.4,14,44 The plaque-activated T-lymphocytes produce INF-?, which inhibits the proliferation of SMC and their ability to synthesize collagen.⁴⁵ The activated macrophages produce metalloproteinases (gelatinase, stromelysine, and interstitial collagenase) that degrade the ECM proteins, favoring the disruption of plaque,⁴⁶ and synthesizing the tissular factor (TF) ,⁴⁷ 1 of the principal activators of the coagulation cascade, which promotes thrombosis of the plaque. These macrophages also induce the apoptosis of the SMC, with a consequent decrease in collagen synthesis and weakening of the fibrous cap, destabilizing the plaque.³⁶

In addition to the monocytes, the T-lymphocytes are equally attracted to the dysfunctional arterial wall by chemoattractant substances, and they are activated at the wall, initiating the production of more cytokines such as INF- γ , TNF- α ; interleukins (IL-1, IL-2, IL-6, IL-8) and growth factors such as GM-CSF that activate the monocytes present in the plaques and favor their proliferation, enabling a local inflammatory response.3,39,40,48

The result of the interaction of these factors is a progression of the atherosclerotic lesion from its initial state of fat striae to complex atherosclerotic plaque.⁴⁹ The rupture or ulceration of the unstable plaque results in the exposure of the procoagulant and prothrombotic surfaces to the blood, causing the activation of platelets and the formation of thrombi, which can trigger clinical complications by occluding the vessel lumen or producing asymptomatic plaque growth.6,50

Therefore, in all the different developmental studies of atherosclerotic plaques, signs of chronic inflammation can be noted, and various physiopathological mechanisms that influence the development, progression, and instability of atherosclerotic lesions have been described.³

INFLAMMATION AND ATHEROSCLEROTIS

Traditionally, atherosclerosis has been considered an illness caused by the accumulation of lipids, where the vulnerable plaques are those with greater lipidic nuclei and a thin fibrous cap, whose rupture would respond to the forces of mechanical stress. Nevertheless, today there is scientific evidence that confirms the role the inflammatory response, whether local or systemic, plays in the development of the atherosclerotic process and in triggering acute cardiovascular events (Figure 1).⁵¹ Patients with unstable angina have elevated reac-

Fig. 2. The local or systemic inflammation leads to the liberation of cytokines, which promotes the synthesis of inflammatory mediators that favor the development of atherosclerosis.

tant values in the acute stage (reactive protein-C [RPC]), serum amyloidal A, fibrinogen) and proinflammatory cytokines (IL-1, IL-6, IL-8)⁵² (Figure 2). The elevated levels of these inflammation inhibitors (RPC, fibrinogen, amyloid A, IL-6) are sensitive mar-

kers for inflammation and correlate with the development of coronary artery disease and its severity,^{53,54} as well as the presence of acute coronary events.^{55,56} The RPC, in addition to being considered an independent predictor of the development of cardiovascular complications, appears to play a role in the pathogenesis of atherosclerosis, as it has been associated with an increase in the risk of thrombosis as it has been associated with an increased risk of cardiovascular complications by promoting the expression of tissular factor by the monocytes 57 and activating the complement cascade. 51 It has also been observed that it promotes the expression of adhesion molecules by the endothelial cells and increases cLDL capture by the macrophages in the plaque by opsonization.⁵⁸

IL-6 is a cytokine with potent pro-inflammatory properties that induces the expression of reactants in the acute phase (the major inducer of hepatic production of RPC) and the migration and differentiation of the activated macrophages.⁵⁹ It also contributes to triggering acute coronary syndromes by enabling the synthesis of metalloproteinases and the expression of LDL receptors in the macrophages, as well as an increase in cLDL capture and the secretion of chemoattractant substances, such as MCP-1, by the same.⁵⁹ Finally, it regulates the expression of adhesion molecules and cytokines, such as IL-1β, and TNF-α, which increases the inflammatory reaction.⁵⁹ At the same time, the liberation of IL-6 is stimulated by IL-1, both acting together and with TNF-α, increasing the synthesis of IL-8 and reactants in the acute phase.⁵²

IL-1 also induces the expression of genes for the synthesis of activating factors of the coagulation system and fibrinolysis inhibitors and the migration of neutrophils to the sub endothelial space, mediated by an increase in the expression of adhesion molecules in the endothelial cells and in the production of GM-CSF (granulocyte and monocyte colony stimulant factor).⁵²

IL-8 ins a pro-inflammatory cytokine produced by various types of cells, including monocytes, macrophages, and T-lymphocytes, and its presence has been detected in frothy cells in human atheroma plaques. It has associated prothrombotic properties by increasing the procoagulant activity of the monocytes, by increasing the synthesis and expression of the tissular factor on the surface of these cells, and pro-atherogenic properties by decreasing MPTI-1 (metalloproteinase tissular inhibitor 1) properties, which favors the predominance of the degradation of the fibrous cap of the plaque over synthesis.⁶⁰

The synthesis of pro-inflammatory cytokines is mediated in large part by the nuclear transcription factor NF-κβ. This is associated with the induction of protein codifying genes, which are vital for the inflammatory processes related to the rupture of atherosclerotic plaques. This factor is activated by diverse stimuli, such as cytokines, viruses, mitogenes, pathogenic microorganisms, modified LDL, oxidative stress, etc.61 The activation of this factor has been detected in macrophages, endothelial cells, and SMC of atherosclerotic plaques,62,63 and there are experimental studies that demonstrated direct correlation between NF-κβ activity and the severity of the coronary lesions.⁵¹ NF- $\kappa\beta$ is found in the form of an inactive heterodimer in the cytoplasm bound to protein inhibitors that are generically known as I $\kappa\beta$.³⁶ This heterdimer consists of 2 sub-units, p50 y p65. When the cell is activated by 1 of the previously mentioned agents, I κβ phosphorilates and undergoes ubiquitination, which acts as a signal for its protelitic degradation. The p50/65 dimer relocates to the nucleus and activates the transcription of target genes that induce the expression of cytokines (TNF- α), interleukins (IL-1, IL-2, IL-6, IL-8), growth factors (MCSF, GM-CSF, G-CSF), chemoattractant substances (MCP-1), adhesion molecules (ICAM-1, VCAM-1, E-selectin), and enzymes (MMP, iNOS, $COX-2$ ^{64,65} that enable the local inflammatory response and destabilize the atherosclerotic plaque.

The inflammatory cells present in the atheromatous plaque express the immune mediator CD40 and its ligand CD40L.⁶⁶ The existence of positive T-cells for accumulated CD40L in the plaques, principally in plaque rapid growth areas and with a greater tendency to complication, suggests that this ligand intervenes in the pathology of the process. The interaction of CD40 with its ligand promotes a humeral and cell response cellular.⁶⁷ The interruption of this union via the administration of anti-CD40L antibodies limits, experimentally, the development of certain autoimmune illnesses such as lupus nephritis, multiple sclerosis, thyroiditis, implant rejection disease, and others.68-72 It has also been shown *in vitro* that the interaction of CD40 and CD40L activates functions related to atherogenesis, including the production of pro-inflammatory cytokines, $\overline{71}$ metalloprteinases, $\overline{73,74}$ and the expression of adhesion molecules⁷⁵ and tissular factor.⁷⁴

THE ROLE OF IL-10 IN ATHEROSCLEROSIS

Among the anti-inflammatory cytokines, IL-10 is considered the anti-inflammatory interleukine *par excelance*. 76,77 It was originally identified as the inhibitory factor in cytokine synthesis (CSIF) ,⁷⁸ as it inhibits the production of cytokines by T-lymphocytes, particularly IFN- γ , by Th1 cells in the murine systems.⁷⁹ Nevertheless, this inhibition is only seen when the macrophages act as antigen presenting cells (APC).⁸⁰

Later studies revealed that IL-10 is in fact a cytokine with pleiotropic properties that acts on different types of cells, including thymocytes, ⁸¹ cytotoxic Tcells,⁸² mastocytes,⁸³ B84 cells, and monocytes-macrophages.⁸⁰

IL-10 is principally produced by a lymphocyte subtype $CD4+$ (Th2) and also in large quantities by macrophages. It is a cytokine with potent anti-inflammatory properties that is capable of inhibiting important functions of these 2 types of cells.76,79 Therefore, it has been described as inhibiting the production of proinflammatory cytokines by macrophages and T85

cells, activated by various stimuli.

IL-10 has been identified in early and advances atherosclerotic lesions, $86,87$ principally located in the cytoplasm of the macrophages, although it is also located in SMC and the extracellular matrix.⁸⁷

Inflammatory properties of IL-10: action mechanisms

One of the first properties attributed to IL-10 was its capacity to inhibit the synthesis of cytokines. There are many published studies supporting this affirmation. Waal Malefyt et al⁷⁹ showed that both human IL-10 or its viral recombinant form added to monocyte cultures activated by INF-γ or LPS, or both, as is produced endogenously in response to these stimuli, was capable of inhibiting the production of inflammatory cytokines, including IL-1α, IL-1β, IL-6, IL-8, TNF-α, INF-γ, GM-CSF and G-CSF by the monocytes. Added to these effects, endogenous IL-10 has an auto regulatory effect on it own production, reducing the synthesis of mRNA IL-10 by the activated monocytes. Wang et al⁸⁵ showed that it could also act on T-cells, inhibiting the production of IL-2, TNF-β, INF-γ and GM-CSF. It also decreased the expression of major histocompatability complex class II molecules (MCH II) by the monocytes/macrophages, and the capacity of these to act as antigen presenting cells, ultimately limiting the specific proliferative antigen response of T-lymphocytes^{79,88} and, therefore, the inflammatory response.

Various mechanisms have been proposed for the manner in which IL-10 inhibits the synthesis of pro-inflammatory cytokines. One of the most studied is the inhibition of the nuclear transcription factor NF-κβ by IL-10 in monocytes and T-cells by the intervention of secondary oxygen free radical messengers.⁷⁹ This results in a reduction in the synthesis of pro-inflammatory interleukins, adhesion molecules, growth factors, and chemoattractants of immunological system cells that limit the local inflammatory response in the plaque. This IL-10 action mechanism differs from that of IL-4, another anti-inflammatory interleukine that also inhibits the synthesis of pro-inflammatory factors, but by a process that does not involve NF-κβ but secondary to an increase in mRNA degradation of said molecule.⁸⁵

O´Farrel et al proposed as another possible mechanism to explain the anti-inflammatory effects of IL-10 the inhibition of interferon production via the activation of STAT group transcription factors.⁸⁹ In a parallel manner, it has been demonstrated that IL-10 can inhibit the expression of pro-inflammatory genes that present areas rich in AU (ARE),⁹⁰ such as TNF- α , IL-1 α , IL-1β, βGM-CSF, IL-8, and others, destabilizing its mRNA by acting on these ARE mofits.⁹¹

In addition, it has been shown that IL-10 is capable of inhibiting monocyte response mediated by CD40CD40L interaction, which appears play relevant role in atherosclerosis.⁶⁷ Mach et al demonstrated that the *in vivo* block of this interaction by antibodies in mice subjected to an atherogenic diet limited the size of the atherosclerotic plaques, reduced it lipidic content, T-cells and macrophages, and decreased the expression of adhesion molecules (VCAM-1).⁶⁷ Therefore, this experimentsl animal model demonstrates another mechanism by which IL-10 appears to play a protective role by limiting the development of atherosclerotic lesions.92,93

IL-10 modulates the immune cell response

We have seen that T-cells participate in the immunological response via the liberation of cytokines. Within the helper T-cells there are 2 types that may mediate different immunological responses. Th1 cells principally produce IL-2 and INF-γ, which is associated with the activation of macrophages and other Tcell subtypes. On the other hand, Th2 cells basically synthesize IL-4 and IL-5, which increase the humeral response and inhibit the Th1 cell response, which predominates in unstable atherosclerotic plaques.⁹⁴

IL-10, together with IL-12, plays an important part in the regulation of these 2 types of immunological response. IL-12 is an important T-cell growth factor⁹⁵ that is principally produced by activated monocytes⁹⁶ and selectively induces a Th197 pattern of immunological response. Uyemura et al⁸¹ showed the presence of $IL-12$ in atherosclerotic plaques. This enables the chronic inflammatory response of the plaque T-cells and macrophages, leading to the destabilization and rupture of the plaque by various mechanisms. Among these is the liberation of INF-γ ⁴⁸ by Th1 cells, stimulating the synthesis of MMP by the macrophages and producing a decrease in the expression of genes for collagen synthesis by SMC and an increase in the apoptosis in said cells, finally destabilizing the plaque by weakening its fibrous cover. In addition to role of regulating the immune cell response, INFγ is capable of potentiating VCAM-1, MHC II, and LDL receptors in vascular cells.⁴⁸

An experimental study on apoloprotein E deficient mice showed that the MRNA of IL-12 is detected early in atherosclerotic plaques that the mRNA of IL-10, and that the daily administration of IL-12 accelerated the development of atherosclerosis in the mice.⁹⁸

The Uyemura group⁸¹ proposed that the endogenous production of IL-10 by T-cells and activated human monocytes, in response to modified LDL stimulation, inhibits IL-12 production and, therefore, facilitates the Th2 immune response decreasing the pro-inflammatory response. These findings suggest that a cross-regulation exists in the production of IL-10 and IL-12 that modulates the local inflammatory response.

The presence of IL-10 in atherosclerotic plaques

IL-10 has been identified in early and advances stages of atherosclerotic lesions, principally located in the macrophage cytoplasm, although it is also in SMC and the extracellular matrix (Figure 1). $86,87$

Mallat et al^{87} showed, in humans, not only the presence of IL-10 in atherosclerotic plaques but also that a strong association exists between the high IL-10 values in the lesions and a reduction in iNOS (nitric oxide sythetase) expression and cell death in the plaques. $99,100$ This suggests that IL-10 plays an important role in limiting the local inflammatory response, preventing excessive cell death in the plaque and promoting, consequently, its stability.

It is known that NO is produced under normal conditions by endothelial cells and that it has important vasodilator and antiatherogenic properties, by inhibiting plaque aggregation, the activation of adhesion molecules, and the proliferation and migration of SMC.¹⁰¹ In initial studies of atherosclerosis, before the lesions visible on angiography appeared, endothelial dysfunction was already present, which caused a reduced NO bioavailability, whether by a decrease in its synthesis, 102 its liberation, 103

or by an increase in its inactivation. ¹⁰⁴ On the other hand, it promotes the synthesis of iNOS, allowing it to bind to super oxidized anions. As a consequence, it produces an increase in the oxidative stress of the atherosclerotic lesion and with it a tendency to vasoconstriction, greater LDL intravascular oxidation, and the activation of NF-kβ, promoting the expression of genes causing the inflammatory response and stabilizing the plaque.¹⁰⁵

Toshiyuki et al¹⁰⁰ showed in *in vitro* studies that endogenous IL-10 plays an essential role in protecting macrophages infected by salmonella from cell death by preventing the excessive production of TNF- α by destroying the bacteria. TNF- α has been shown to be capable of inducing cell death by apoptosis in various types of cells, in addition to being and important mediator of inflammatory effects.^{106,107}

Cohen et al⁹⁹ later showed that *in vitro* IL-10 could inhibit the apoptosis of T-cells, and that this was mediated in part by the over-expression of Bcl-2 protein, preserving in the lymphocytes rescure from apoptosis their capacity to proliferate upon IL-2 stimulation.

Interleukin-10 and the vulnerability of atherosclerotic plaque

It is known that one of the principal conditions for the destabilization of atherosclerotic plaques is the degradation of the extraxcellular matrix (ECM) and the fibrous collagen cover. The macrophages present in the plaques are in charge of modulating the exchange

of the ECM, synthesizing and segregating degrading ECM enzymes, called metallproteinases (MMP), as well as its corresponding inhibitors (MPTI). The principal MMP studied are interstitial collagenase, stromelysine, and gelatinase 92 Kid and 72Kd. These are initially synthesized inactively and are later activated by various stimuli such as oxidative stress, and pro-inflammatory cytokines.¹⁰⁸ There are also 2 types of MMP inhibitors: MPTI-1 and MPTI-2.¹⁰⁹ MPTI-1 interacts with active forms of collagenase and stromelysine, as well as with the precursor and active form of gelatinase 92Kd. MPTI-2 specifically inhibits the proenzyme and active form of gelatinase 72Kd.¹⁰⁸

Lacraz et al¹⁰⁸ showed in *in vitro* experiments that IL-10 has a specific regulatory effect on macrophag4es and monocytes, overriding MMP synthesis and stimulating instead the synthesis of its inhibitor, MPTI-1. This suggests that IL-10 has powerful anti-inflammatory effects, counteracting the degradation of the macrophages and changing the protease/anti-protease balance, favoring the preservation of EMV and the fibrous cover, giving the plaque stability.

Regulation of prothrombotic events: the role of IL-10

One of the important factors in the triggering of an acute coronary syndrome is arterial occlusion by a thrombus superimposed on a complicated plaque.3,39,49,110,111 Among the triggers of intravascular thrombosis is the expression by endothelial cells and monocytes of the tissular factor (TF).¹¹²

This is 1 of the principal initiators of the coagulation cascade *in vivo*, by binding to factor VII and favoring its activation. The complex formed by TF/FVIIa later activates factors X and IX from the common coagulation pathway.

TF is not expressed in the cells under normal conditions, but as a response to various stimuli, endotoxine (LPS) being the most effective, but also to pro-inflammatory cytokines (IL-1, MCP-1, growth factor from PDGF platelets, and others).¹¹³

In 1993, Pradier et al¹¹⁴ showed in *in vitro* experiments on isolated monocytes that IL-10 has and inhibitory effect on the expression of TF by the cells in response to the previously mentioned stimuli, that is transcriptional inhibition (mRNA).¹¹⁵

These findings were later confirmed by Landmark et al,¹¹⁶ who observed that IL-10 maintained its inhibitory effect *in vivo* over TF expression by the monocytes when induced by LPS, also acting on mRNA. In another recent study on humans in whom endotoxemia was experimentally induced, IL-10 was shown to be capable of inhibiting the action of the coagulation system as well as attenuating fibrinolysis.¹¹⁷

All the preceding suggests that IL-10 could be very

useful in the treatment of some pathologies that present an increased risk of thrombosis due to monocyte procoagulant activity, such as DIC or ischemic heart disease.¹¹⁶

Experimental evidence of the protective role of IL-10 in atherosclerosis

IL-10 is cytokine that has an important regulatory effect on the immune response. Its capacity to inhibit the synthesis of cytokines and various cell functions of macrophages and T-lymphocytes converts IL-10 into a powerful anti-inflammatory agent. If we extrapolate this concept to atherogenesis, understood to be a chronic inflammatory disease of the vascular wall, we can hypothesize that this molecule could play a protective role in the pathogenesis of atherosclerosis.

Multiple experimental *in vivo* and *in vitro* animal studies have been performed, and the results support the theory of the protective role of IL-10, in the formation as well as in the stabilization of atherosclerotic plaque. Mallat et al¹¹⁸ demonstrated that C573L/6J mice deficient in IL-10 (IL 10-/-) had increased susceptibility to the development of atherosclerotic lesions compared to wild mice (producers of IL-10, IL 10+/+). Also, on studying the composition of atherosclerotic plaques, it was observed that in the IL10-/-, mice there was a greater infiltration of inflammatory cells, increased IFN-γ (characteristic of the Th1 response) and a lower collagen content with respect the plaques of wild mice, findings that suggest that these plaques are more vulnerable or unstable with a higher tendency to rupture.³ Later, on studying the effect of DNA transfer of IL-10 to the IL-10-/- mice fed an atherogenic diet they achieved a 60% reduction in the size of the atherosclerotic lesions. On the other hand, they proved that in IL-10-/- mice in an environment free of pathogens, the total atherosclerotic lesion surface was 4 to 5 times less than the IL-10-/- mice subjected to normal conditions, in spite of not finding differences in their lipidic profile. This data supports the theory of the intervention of pathogenic microorganisms in the development of atherosclerosis.¹¹⁹

In summary, this study demonstrates that IL-10 has a profound impact both on the development and the composition of atherosclerotic lesions, as well as a protective effect against environmental pathogen. These findings were later corroborated by Pinderski et al,¹²⁰ who demonstrated *in vivo* that transgenic IL-10 mice over-expressing IL-10 in T-cells, upon being fed an atherogenic diet showed a significant reduction in the development of atherosclerotic lesions as compared to wild mice or IL-10 deficient mice under the same conditions (Figure 3). It was also observed that the latter also developed larger lesions, with more inflammatory infiltrate, lipidic infiltrate, and a nearly imperceptible fibrous cover. In a parallel manner, the

experiments *in vitro* of this group showed that pretreatment with IL-10 in these mice could inhibit the interaction of LDL-activated monocytes with the endothelium and their adhesion to same. This may be explained in part by the capability of IL-10 to inhibit the expression of adhesion molecules (VCAM-1, ICAM-1) by endothelial cells.

Similar results have been obtained in a recent study performed in humans that supports the hypothesis of the protective role of IL-10 in atherogenesis by contributing to the maintenance of plaque stability, avoiding acute events. Smith et $al¹²¹$ demonstrated that patients with unstable angina had significantly lower amounts of IL-10 in their blood than those with chronic stable angina, suggesting that low IL-10 values are associated with greater clinical stability (Figure 4).

On the other hand, a significant liberation of IL-10 has been shown in multiple studies of myocardial ischemia-reperfusion and pulmonary bypass in humans.¹²²⁻¹²⁴ Yang et al¹²⁵ observed in an experimental model of ischemia-reperfusion in IL 10-/- mice that they had an exaggerated inflammatory response in the reperfused tissues as compared to wild mice. This was revealed as an increase in neutrophil infiltration in the reperfused tissues and an increase in production of TNF-α, ICAM-1 and products of NO degradation, ultimately leading to an increase in the size of the AMI and myocardial necrosis as well as the mortality rate in the immundepressed mice.

These findings provide evidence that the endogenous production of IL-10 has protective effects against myocardial infarction and reperfusion damage by inhibiting the production of TNF- α , iNOS, the expression of adhesion molecules, and recruitment of neutrophils. There are currently many studies underway that suggest the potential use of IL-10 in various case of reperfusion damage.¹²⁶⁻¹³¹

Other properties of IL-10: a new therapeutic agent?

Recently, anti-tumor properties have been associated with IL-10 related to its capability of decreasing the synthesis of VEGF (vascular endothelial growth factor), TNF-α and MMP-9 (92Kd gelatinase 92Kd), as well as in preventing the angiogenesis associated with tumor growth. $132,133$ The main stimulus for angiogenesis is ischemia and inflammation, and both conditions are found narrowly related to ischemic illness.134,135

Given the anti-inflammatory properties attributed to IL-10, it is being studied as possible therapy for a great number of chronic diseases, including rheumatoid arthritis, inflammatory intestinal disease, multiple sclerosis, eosinophilic allergy, Wegener granulomatosis, cardiac transplant rejection, and others.¹³⁶ For example, the pathogenesis of inflammatory intestinal disease (Crohn´s disease and ulcerative colitis) is characterized by an unbalance in the activity of Th1 and Th2 lymphocytes, with a predominantly Th1 immune response, that cause a massive inflammatory response in the intestinal mucous. IL-10 is an anti-inflammatory cytokine that regulates the production of pro-inflammatory cytokines derived from Th1 lymphocytes, promoting a Th2 immune response, which is essential in the fight against inflammation.¹³⁷ The topical, intravenous, and subcutaneous use of recombinant human IL-10 (rhuIL-10) has been studied in humans with inflammatory intestinal disease.¹³⁸ While the results obtained by Fedorak et al¹³⁸ promoted the use of IL-10 by finding a clinical and endoscopic improvement in illness after the subcutaneous administration of rhuIL-10 in patients with Crohn's disease, Colombel et al¹³⁹ were not able to demonstrate that the treatment prevented the endoscopic recurrence of the disease in post-operative patients with Crohn´s disease. The discrepancies in these results may be due to the difficulty in selecting a subgroup of patients who would potentially benefit from the biological therapy, as well as identifying the most appropriate way to administer the required $dose.¹⁴⁰$

It was observed in experimental animal studies that IL-10 deficient mice spontaneously develop a severe form of enterocolitis.¹⁴¹ Nevertheless, the transfer of CD4+ T-cells from transgenic IL-10 mice (which over express IL-10) is capable of overcoming, in IL10-/ mice, the development of colitis¹⁴² in situations in which it would normally occur.

The results of the studies mentioned suggest that IL-10 should be considered a new therapeutic tool in the field of atherosclerosis treatment. Nevertheless, the administration of IL-10 as long-term therapy, with all its immunosupressor actions, may present unexpected consequences as it is able to potentially produce an antigen-specific energy. Different experimental studies have shown that IL-10 increases susceptibility to certain infections, principally those involving intracellular pathogens such as *Chlamydia* and *Listeria monocytogenes.*¹³⁶ The recombinant BALB/c mice stimulated with *Chlamydia* produced higher IL-10 values than the C57BL/6J wild mice. The BALB/c mice presented, as a consequence, a less aggressive inflammatory response against the *Chlamydia* infection, succumbing in a greater number than the C57BL/6J143 mice.

On the other hand, the IL 10-/- mice, deficient in IL-10, upon being infected with subtle doses of *Chlamydia*, developed less granulomatous lesions than wild mice.¹⁴⁴

CONCLUSIONS

The current knowledge about the physiopathology of atherosclerosis has changed radically from that of the last decades. Today it is generally accepted that inflammation plays a fundamental role in the develop-

Fig. 3. Surface of the atherosclerotic lesion in the aortic root according to serum IL-10 values in wild mice (control), transgenic mice (IL10+/+), and IL-10-deficient mice (IL-10-/-). Observe the smaller surface of the lesion in the transgenic mice vs the controls (5.433±4.008 vs 13.574±4.212 mm² ; *P<.05). Note the marked increase in the lesion size of the IL-10-/- deficient mice vs the IL-10+/+ mice (33.250±9.117 mm²; *P<.0001) (modified from Pinderski O, et al^{120}

Fig. 4. Mean serum concentrations of IL-10 (pg/ml) in patients with unstable and chronic unstable angina (14±10.1 vs 28.43+12.1; P <.0001) (modified from Smith D, et al,¹²¹).

ment and progression of atherosclerotic lesions, leading in the long or short term to the appearance of clinical signs. Nevertheless, the intrinsic mechanism by which this inflammatory response is triggered and develops continues to be clearly understood. The best knowledge of the physiopathological phenomenon underlying the process of atherogenesis will allow new investigative paths to be opened to combat the disease.

From this point of view, various studies performed to study the relevance of IL-10 in atherosclerosis suggest that it has a protective role limiting the local inflammatory response, which favors the progression and instability of the atherosclerotic plaque, eventually leading to the development of acute coronary syndromes. This allows us to postulate the possible role of IL-10 as a therapeutic agent whose exogenous administration restrains the development of lesions and confers stability, improving the clinical course of the patient. IL-10 could also be a new risk marker that allows us to predict plaque instability and its propensity toward complications.

REFERENCES

- 1. Breslow JL. Cardiovascular disease burden increases, NIH funding decreases. Nat Med 1997;3:600-1.
- 2. Braunwald E. Shattuck Lecture-cardiovascular medicine at the turn of the millennium: trumps, corcerns, and opportunities. N Engl J Med 1997;337:1360-9.
- 3. Ross R. Atherosclerosis- an inflammatory disease. N Engl J Med 1999;340:115-26.
- 4. Moreno PR, Fallon Jt. Inflammation in acute coronary syndromes. En: Schultheiss H, Schwimmbeck P, editors. The role of immune mechanism in cardiovascular disease. Berlin: Springer, 1997; p. 213-29.
- 5. Van der Wal AC, Becker AE, van der Loss CM, Das PK. Site of intimal rupture or erosion af thrombosed coronary atherosclerotic plaques is characterized by an inflammatory process irrespective of the dominant plaque morphology. Circulation 1994;89:36-44.
- 6. Davies MJ. Stability and instability: the two faces of coronary atherosclerosis: The Paul Dudley White Lecture, 1995. Circulation 1996;93:1354-63.
- 7. Braunwald E, Fuster V. Unstable angina. Definition, pathogenesis, and classification. En: Fuster V, Toss R, Topol EJ, editors. Atherosclerosis and coronary disease. Philadelphia: JB Lippincot, 1996; p. 1285-98.
- 8. Ridker PM, Cushman M, Stamper MJ, Tracy RP, Hennekens CH. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. N Engl J Med 1997;336:973-9.
- 9. Hasson GK, Holm J, Kral JG. Accumulation of IgG and complement factor C3 in human arterial endothelium and atherosclerotic lesions. Acta Pathol Microbiol Inmunol Scand 1984;92A: 429-35.
- 10. Roselaar SE, Schonfeld G, Daugherty A. Enhanced development of atherosclerosis in cholesterol-fed rabbits by suppression of cell-mediated inmunity. J Clin Invest 1995;96:389-94.
- 11. Emeson EE, Shen ML. Accelerated atherosclerotic in hyperlipidemic C57BL/6 mice treated with cyclosporin A. Am J Pathol 1993;142:1906-15.
- 12. Fyfe AI, Qiao J-H, Lusis A. Inmune-deficient mice develop typical atherosclerotic fatty streaks when fed an atherogenic diet. J Clin Invest 1994;94:2516-20.
- 13. Barath P, Fishlein MC, Cao J, Berenson J, Helfant RH, Forrester JS. Detection and localization of tumor necrosis factor in human atheroma. Am J Cardiol 1990;65:297-302.
- 14. Clinton SK, Fleet JC, Loppnow H, Salomon RN, Clark BD, Cannon JG, et al. Interleukin-1 gene expression in rabbit vascular tissue in vivo. Am J Pathol 1991;138:1005-14.
- 15. Seino Y, Ikeda U, Ikeda M, Yamamoto K, Misawa Y, Hasegawa T, et al. Interleukin-6 gene transcripts are expressed in human atherosclerotic lesions. Cytokine 1994;6:87-91.
- 16. Kishikawa H, Shimokama T, Watanabe T. Localization of T lymphocytes and macrofages expressing IL-1, IL-2 receptor, IL-6 and TNF in human aortic intima. Role of cell-mediated immunity in human atherosclerosis. Virchows Arch 1993;423:433-42.
- 17. Lee RT, Libby P. The unstable atheroma. Arterioscler Thromb

Vasc Biol 1997;17:1859-67.

- 18. Suwaidi JA, Hamasaki S, Higano ST, Nishimura RA, Holmes DR Jr, Lerman A. Long-term follow-up of patients with mild coronary disease and endothelial dysfunction. Circulation 2000;101:948-54.
- 19. Zeiher AM, Drexeler H, Wollschlyger H, Just H. Modulation of coronary vascular tone in humans: progressive endothelial dysfunction with different early stages of coronary atherosclerosis. Circulation 1991;83:391-401.
- 20. Quyyumi AA. Endothelial function in health and disease: new insights into the genesis of cardiovascular disease. Am J Med 1998;105:32S-9S.
- 21. Lüscher TF, Vanhoutte PM. The Endothelium: modulator of cardiovascular function. Boca Raton: CRC Press, 1990.
- 22. Moncada S, Vane JR. Arachidonic acid metabolites and the interactions between platelets and blood-vessel walls. N Engl J Med 1979;300:1142-7.
- 23. Vane JR, Anggard EE, Botting RM. Regulatory functions of the vascular endothelium. N Engl J Med 1990;323:27-36.
- 24. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature 1980;288:373-6.
- 25. Liao JK. Endothelium and acute coronary syndromes. Clin Chem 1998;44:1799-808.
- 26. Drexler H. Endothelium dysfunction: clinical implications. Prog Cardiovasc Dis 1997;4:287-324.
- 27. Simon A, Castro A, Kaski JC. Avances en el conocimiento de la disfunción endotelial y su aplicación en la práctica clínica. Rev Esp Cardiol 2001;54:211-7.
- 28. Ohara Y, Peterson TE, Harrison DG. Hypercholesterolemia increases endothelial superoxide anion production. J Clin Invest 1993;9:2546-51.
- 29. Vogel RA. Cholesterol lowering and endothelial function. Am J Med 1999;107:479-87.
- 30. Perticone F, Ceravolo R, Pujia A, Ventura G, Iacopino S, Scozzafava A, et al. Prognostic significance of endothelial dysfunction in hypertensive patients. Circulation 2001;104:191-6.
- 31. Cooke JP, Tsao RS. Is NO an endogenous antiatherogenic molecule? Artherioscler Thromb 1994;14:653-5.
- 32. Sánchez-Recalde A, Kaski JC. Diabetes mellitus, inflamación y aterosclerosis coronaria: perspectiva actual y futura. Rev Esp Cardiol 2001;54:751-63.
- 33. Klatt P, Esterbauer H. Oxidative hypothesis of atherogenesis. J Cardiovasc Risk 1996;3:346-51.
- 34. Berliner JA, Navab M, Fogelman AM, Frank JS, Demer LL, Edwards PA, et al. Atherosclerosis: basic mechanisms. Oxidation, inflammation, and genetics. Circulation 1995;91:2488-96.
- 35. Jang Y, Lincoff AM, Plow EF, Topol EJ. Cell adhesion molecules in coronary disease. J Am Coll Cardiol 1994;24:1591-601.
- 36. Martínez-González J, Llorente-Cortes V, Bandimon L. Biología celular y molecular de las lesiones ateroscleróticas. Rev Esp Cardiol 2001;54:218-31.
- 37. Liao JK, Shin WS, Lee WY, Clark SL. Oxidazed low-density lipoprotein decreases the expression of endothelial nitric oxide synthase. J Biol Chem 1995;270:319-24.
- 38. Sata M, Walsh K. Oxidized LDL activates Fas-mediated endothelial cell apoptosis. J Clin Invest 1998;102:1682-9.
- 39. Libby P. The molecular bases of the acute coronary syndromes. Circulation 1995;91:2844-50.
- 40. Jonassson L, Holm J, Skalli O, Bondjers G, Hansson GK. Regional accumulations of T cells, macrophages, and smooth muscle cells in the human atherosclerotic plaques. Arteriosclerosis 1986;6:131-8.
- 41. Freeman MW. Macrophage scavenger receptors. Curr Opin Lipid 1994;5:143-8.
- 42. Weissberg PL. Atherogenesis: current understanding of the causes of atheroma. Heart 2000;83:247-52.
- 43. Libby P. Current concepts of the pathogenesis of the acute coronary syndromes. Circulation 2001;104:365-72.
- 44. Falk E, Shah PK, Fuster V. Coronary plaque disruption. Circulation 1995;92:657-71.
- 45. Serneri GGN, Abbate R, Gori AM, Attanasio M, Martini F, Giusti B, et al. Transient intermittent lymphocyte activation is responsible for the instability of angina. Circulation 1992;86: 790-7.
- 46. Shah PK, Falk E, Badimon JJ, Fernández-Ortiz A, Mailhac A, Villareal-Levy G, et al. Human monocyte-derived macrophages induce collagen breakdown in fibrous cap of atherosclerotic plaques: potential roleof matrix-degrading metalloproteinases and implications for plaque rupture. Circulation 1995;92: 1565-9.
- 47. Mach F, Schönbeck U, Bonnefoy-Y, Pober JS, Libby P. Activation of monocyte/macrophage functions related to acute atheroma complication by ligation of CD40: induction of collagenase, stromelysin, and tissue factor. Circulation 1997;96:369.
- 48. Hansson GK, Holm J, Jonasson L. Detection of activated T lymphocytes in the human atherosclerotic plaque. Am J Pathol 1989;135:169-75.
- 49. Ross R. The pathogenesis of the atherosclerosis- an update. N Engl J Med 1986;314:488-500.
- 50. Badimon L, Chesebro JH, Badimon JJ. Thrombus formation on ruptured atherosclerotic plaques and rethrombosis on evolving thrombi. Circulation 1992;86(Suppl III):74-85.
- 51. Kaski JC. Inflamación, infección y enfermedad coronaria: mitos y realidades. Rev Esp Cardiol 2000;53:1311-7.
- 52. Biasucci LM, Liuzzo G, Fantuzzi G, Caligiuri G, Rebuzzi AG, Ginnetti F, et al. Increasing levels of interleukin (IL)-1Ra and IL-6 during the first 2 days of hospitalization in unstable angina are associated with increased risk of in-hospital coronary events. Circulation 1999;99:2079-84.
- 53. Mori T, Sasaki J, Kawaguchi H, Handa K, Takada Y, Matsunaga A, et al. Serum glycoproteins and severity of coronary atherosclerosis. Am Heart J 1995;12:234-8.
- 54. Liuzzo G, Biasucci LM, Gallimore JR, Grillo RL, Rebuzzi AG, Pepys MB, et al. The prognostic value of C-reactive protein and serum amyloid-A in severe unstable angina. N Engl J Med 1994;331:417-24.
- 55. Ridker PM, Cushman M, Stampfer MJ, Tracy RP, Hennekens CH. Inflammation, aspirin and the risk of cardiovascular disease in apparently healthy men. N Engl J Med 1997;336:973-9.
- 56. Kuller LH, Tracy Rp, Shaten J, Meilahn EN. Relation of C-reactive protein and coronary heart disease in the MRFIT nested case-control study. Am J Epidemiol 1996;144:537-47.
- 57. Abdelmouttaleb I, Danchin N, Ilardo C, Aimone-Gastin I, Angioi M, Loznievski A, et al. C-reactive protein and coronary artery disease: additional evidence of the implication of an inflammatory process in acute coronary syndromes. Am Heart J 1999;137:346-51.
- 58. Zwaka TP, Hombach V, Torrzewski J. C-reactive protein low density lipoprotein uptake by macrophages. Circulation 2001;103:1194-7.
- 59. Schieffer B, Schieffer E, Hilfiker-Kleiner D, Hilfiker A, Kovanen PT, Kaartinen M, et al. Expression of angiotensin II and interleukin 6 in human coronary atherosclerotic plaques. Potential implications for inflammation and plaque instability. Circulation 2000;101:1372-8.
- 60. Moreau M, Brocheriou I, Petit L, Ninio E, Chapman MJ, Rouis M. Interleukin-8 mediates downregulation of tissue factor inhibitor of metalloproteinase-1 expression in cholesterol-loaded human macrophages. Relevance to stability of atherosclerotic plaque. Circulation 1999;99:420-6.
- 61. Barnes PJ, Karin M. Nuclear Factor-κβ. A pivotal transcription factor in chronic inflammatory diseases. N Engl J Med 1997;336:1066-71.
- 62. Ritchie ME. Nuclear Factor-κβ is selectively and markedly activated in humans with unstable angina pectoris. Circulation 1998;98:1707-13.
- 63. Brand K, Page S, Rogler G, Bartsch A, Brandl R, Knuechel R, et al. Activated transcription factor nuclear-kappa β is present in the atherosclerotic lesion. J Clin Invest 1996;97:1715-2.
- 64. Baeuerle PA, Henkel T. Function and activation of NF-κβ in the immune system. Annu Rev Immunol 1994;12:141-79.
- 65. Bourcier T, Sukhova G, Libby P. The nuclear factor κβ signaling pathway participates in dysregulation of vascular smooth

muscles cells in vitro and in human atherosclerosis. J Biol Chem 1997;272:15817-24.

- 66. Mach F, Schonbeck U, Sukhova GK, Bourcier T, Bonnefoy JY, Pober JS, et al. Functional CD40 ligand is expressed on human vascular endothelial cells, smooth muscle cells, and macrophages: implications for CD40-CD40 ligand signaling in atherosclerosis. Proc Nat Acad Sci 1997;94:1931-6.
- 67. Mach F, Schonbeck U, Sukhova GK, Atkinson E, Libby P. Reduction of atherosclerosis in mice by inhibition of CD40 signalling. Nature 1998;394:200-3.
- 68. Durie FH, Fava RA, Foy TM, Aruffo A, Ledbetter JA, Noelle RJ. Prevention of collagen-induced arthritis with antibody to gp39, the ligand for CD40. Science 1993;261:1328-30.
- 69. Mohan C, Shi Y, Laman JD, Datta SK. Interaction between CD40 and its ligand gp39 in the development of murine lupus nephritis. J Immunol 1995;154:1470-80.
- 70. Larsen CP, Elwood ET, Alexander DZ, Ritchie SC, Hendrix R, Tucker-Burden C, et al. Long-term acceptance of skin and cardiac allografts after blocking CD40 and CD28 pathways. Nature 1996;381:434-8.
- 71. Gerritse K, Laman JD, Noelle RJ, Aruffo A, Ledbetter JA, Boersma WJ, et al. CD40-CD40 ligand interactions in experimental allergic encephalomyelitis and multiple sclerosis. Proc Natl Acad Sci 1996;93:2499-504.
- 72. Carayanniotis G, Master SR, Noelle RJ. Supression of murine thyroiditis via blockade of the CD40-CD40L interaction. Immunity 1997;90:421-6.
- 73. Schonbeck U, Mach F, Sukhova GK, Murphy C, Bonnefoy JY, Fabunmi RP, et al. Regulation of matrix metalloproteinase expression in human vascular smooth muscle cells by T lymphocytes: a role for CD40 signaling in plaque rupture. Circ Res 1997;81:448-54.
- 74. Mach F, Schönbeck U, Bonnefoy J-Y, Pober JS, Libby P. Activation of monocyte/macrophage functions related to acute atheroma complication by ligation of CD40. Induction of collagenase, stromelysin, and tissue factor. Circulation 1997;96: 396- 9.
- 75. Karmann K, Hughes CC, Schechner J, Fanslow WC, Pober JS. CD40 on human endothelial cells: inductibility by cytokines and functional regulation of adhesion molecule expression. Proc Natl Acad Sci 1995;92:4342-6.
- 76. De Vries JE. Immunosupressive and anti-inflammatory properties of interleukin-10. Ann Med 1995;27:537-41.
- 77. Moore K, O'Garra A, De Waal Malefyt R, Vieira P, Mosmann TR. Interleukin-10. Ann Rev Immunol 1993;11:165-90.
- 78. Fiorentino DF, Bond MW, Mosmann TR. Two types of mouse T helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. J Exp Med 1989;170:2081-95.
- 79. De Waal Malefyt R, Abrams RJ, Bennett B, Figdor CG, De Vries JE. Interleukin (IL)-10 inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. J Exp Med 1991;174:1209-20.
- 80. Fiorentino DF, Zlotnik A, Vieira P, Mosmann TR, Howard M, Moore KW, et al. IL-10 acts on the antigen presenting cell to inhibit cytokine production by Th1 cells. J Immunol 1991; 146:3444-51.
- 81. MacNeil IA, Suda T, Moore KW, Mosmann TR, Zlotnik A. Il-10 a novel growth cofactor for mature and inmature t cells. J Immunol 1990;145:4167-73.
- 82. Chen WF, Zlotnik A. IL-10: a novel cytotoxic cell differentiation factor. J Immnol 1991;147:528-34.
- 83. Thompson-Snipes L, Dhar V, Bond MW, Mosmann TR, Moore KW, Rennick DM. Interleukin 10: a novel stimulatory factor for mast cells and their progenitors. J Exp Med 1991;173:507- 10.
- 84. Go NF, Castle BE, Barrett R, Kastelein R, Dang W, Mosmann TR, et al. Interleukin 10, a novel B cell stimulatory factor: unresponsiveness of X chromosome linked immunodeficiency B cells. J Exp Med 1990;172:1625-31.
- 85. Wang P, Wu P, Siegel MI, Egan RW, Billah MM. Interleukin (IL)10 inhibits nuclear factor κβ (NF-κβ) activation in human monocytes: IL-10 and IL-4 suppress cytokine synthesis by diffe-

rent mechanisms. J Biol Chem 1995;270:9558-63.

- 86. Uyemura K, Demer LL, Castle SC, Jullien D, Berliner JA, Gately MK, et al. Cross regulation roles of interleukin (IL)-12 and IL-10 in the atherosclerosis. J Clin Invest 1996;97:2130-8.
- 87. Mallat Z, Heymes C, Ohan J, Faggin E, Leseche G, Tedgui A. Expression of interleukin-10 in advanced human atherosclerotic plaques. Arterioscler Thromb Vasc Biol 1999;19:611-6.
- 88. De Waal Malefyt R, Haanen J, Spits H, Roncarolo MG, Te Velde A, Figdor C, et al. Interleukin 10 (IL-10) and viral IL-10 strongly reduce antigenic-specific human T cell proliferation by disminishing the antigenic-presenting capacity of monocytes via downregulation of class II major histocompatibility complex expression. J Exp Med 1991;174:915-24.
- 89. O'Farrell AM, Liu Y, Moore KW, Mui AL. IL-10 inhibits macrophage activation and proliferation by distinct signaling mechanisms: evidence for Stat3-dependent and –independent pathways. EMBO J 1998;17:1006-8.
- 90. Shaw G, Kamen R. A conserved AU sequence from the 3' unstranslated region of GM-CSF mRNA mediates selective mRNA degradation. Cell 1986;46:659-67.
- 91. Kishore R, Tebo JM, Kolosov M, Hamilton TA. Cutting edge: clustered AU-rich elements are the target of IL-10 mediated mRNA destabilization in mouse macrophages. J Immunol 1999;19:734-42.
- 92. Suttles J, Milhorn DM, Miller RW, Poe JC, Wahl LM, Stout RD. CD40 signaling monocyte inflammatory cytokine synthesis through an ERK1/2-dependent pathway: a target of interleukin 4 (IL-4) and IL-10 anti-inflammatory action. J Biol Chem 1999;274:5835-42.
- 93. Poe JC, Wagner DH, Miller RW, Stout RD, Suttles J. IL-4 and IL-10 modulation of CD40-mediated signaling of monocyte IL-1beta synthesis and rescue from apoptosis. J Immunol 1997;59:846-52.
- 94. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clones I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol 1986;136:2348-57.
- 95. Gately MK, Desai BB, Wolitsky AG, Quinn PM, Dwyer CM, Podlaski FJ, et al. Regulation of human lymphocyte proliferation by a heterodimeric cytokine, IL-12 (cytotoxic lymphocyte maturation factor). J Immunol 1991;147:874-82.
- 96. D'Andrea A, Rengaraju M, Valiante NM, Chehimi J, Kubin M, Aste M, et al. Production of natural killer cell stimulatory cell factor (interleukin 12) by peripheral blood mononuclear cells. J Exp Med 1992;176:1387-98.
- 97. Germann T, Gately MK, Schoenhaut DS, Lohoff M, Mattner F, Fischer S, et al. Interleukin-12/T-cell stimulating factor, a cytokine with multiple effects on T helper type 1 (Th1) but not Th2 cells. Eur J Immunol 1993;23:1762-70.
- 98. Lee TS, Yen HC, Pan CC, Chau LY. Role of interleukin 12 in the development of atherosclerosis in apo-E deficient mice. Arterioscler Thromb Vasc Biol 1999;19:734-42.
- 99. Cohen SB, Crawley JB, Kahan MC, Feldmann M, Foxwell BM. Interleukin-10 rescues T cells from apoptotic cell death: association with upregulation of Bcl-2. Immunology 1997;17: 145-50.
- 100. Arai T, Hiromatsu K, Nishimura H, Kimura Y, Kobayashi N, Ishida H, et al. Endogenous interleukin 10 prevents apoptosis in macrophages during Salmonella infection. Biochem Biophys Res Commun 1995;213:600-7.
- 101. Moncada S, Palmer RM, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. Pharmacol Rev 1991;43: 109-42.
- 102. Verbeuren TJ, Coene MC, Jordaens FH, Van Hove CE, Zonnekeyn LL, Herman AG. Effect of hypercholesterolemia on vascular reactivity in the rabbit. Circ Res 1986;589:496-504.
- 103. Shimokawa H, Vanhoutte PM. Impaired endothelium-dependent relaxation to aggregating platelets and related vasoactive substances in porcine coronary arteries in hypercholesterolemia and atherosclerosis. Circ Res 1989;64:900-14.
- 104. Miyazaki H, Matsuoka H, Cooke JP, Usui M, Ueda S, Okuda S, et al. Endogenous nitric oxide synthase inhibitor. A novel marker of atherosclerosis. Circulation 1999;99:1141-6.
- 105. Gurfinkel E. Infección y aterosclerosis. Rev Esp Cardiol

2001;54:383-92.

- 106. Cohen JJ, Duke RC, Fadok VA, Sellins KS. Apoptosis and programmed cell death in immunity. Annu Rev Immunol 1992; 10:267-93.
- 107. Beutler B, Cerami A. The biology of cachectin/TNF-a primary mediator of the host response. Annu Rev Immunol 1989;7:625-55.
- 108. Lacraz S, Nicod LP, Chicheportiche R, Welgus HG, Dayer JM. IL-10 inhibits metalloproteinase and stimulates TIMP-1 production in human mononuclear phagocytes. J Clin Invest 1995;96:2304-10.
- 109. Albin RJ, Senior RM, Welgus HG, Connolly NL, Campbell EJ. Human alveolar macrophages release an inhibitor of metalloproteinase elastase in vitro. Am Rev Respir 1987;135:1281-5.
- 110. Fuster V, Badimon L, Badimon JJ, Chesebro JH. The pathogenesis of coronary artery disease and the acute coronary syndromes (Part I). N Engl J Med 1992;326:242-50.
- 111. Fuster V, Badimon L, Badimon JJ, Chesebro JH. The pathogenesis of coronary artery disease and the acute coronary syndromes (Part II). N Engl J Med 1992;326:310-8.
- 112. Camerer E, Kolsto A, Prydz H. Cell biology of tissue factor, the principal initiator of blood coagulation. Thrombosis Research 1996;81:1-41.
- 113. Ernofsson M, Siegbahn A. Platelet-derived growth factor and monocyte chemostatic protein-1 induce human peripherical blood monocytes to express tissue factor. Thrombosis Research 1996;81:307-20.
- 114. Pradier O, Gerard C, Delvaux A, Lybin M, Abramowicz D, Capel P, et al. Interleukin-10 inhibits the induction of monocyte procoagulant activity by bacterial lipopolyshacharide. Eur J Immunol 1993;23:2700-3.
- 115. Ramani M, Ollivier V, Khechai F, Vu T, Ternisien C, Bridey F, et al. Interleukin-10 inhibits endotoxin-induced tissue factor mRNA production by human monocytes. FEBS Letters 1993a;334:114-6.
- 116. Lindmark E, Tenno T, Chen J, Siegbahn A. Il-10 inhibits LPSinduced human monocyte tissue factor expression in whole blood. Br J Haematol 1998;102:597-604.
- 117. Parjkt D, van der Poll T, Levi M, Cutler DL, Affrime MB, Van den Ende A, et al. Interleukin-10 inhibits activation of coagulation and fibrinolysis during human endotoxemia. Blood 1997;89:2701-5.
- 118. Mallat Z, Besnard S, Duriez M, Deleuze V, Emmanuel F, Bureau MF, et al. Protective role of interleukin-10 in atherosclerosis. Circ Res 1999;85:E17-E24.
- 119. Libby P, Egan D, Skarlattos S. Roles of infection agents in atherosclerosis and restenosis: an assessment of the evidence and need for future research. Circulation 1997;96:4095-103.
- 120. Pinderski Oslund LJ, Hedrick CC, Olvera T, Hagenbaugh A, Territo M, Berliner JA, et al. Interleukin-10 blocks atherosclerotic events in vitro and in vivo. Arterioscler Thromb Vasc Biol 1999;19:2847-53.
- 121. Smith D, Irving S, Sheldon J, Cole D, Kaski JC. Serum levels of the antiinflammatory cytokine interleukin-10 are decreased in patients with unstable angina. Circulation 2001;104:746-9.
- 122. Seghaye M, Duchataeu J, Bruniaux J, Demontoux S, Bosson C, Serraf A, et al. Interleukin-10 release related to cardiopulmonary bypass in infants undergoing cardiac operations. J Thorac Cardiovasc Surg 1996;111:545-53.
- 123. Wan S, Marchant A, DeSmet JM, Antoine M, Zhang H, Vachiery JL, et al. Human cytokine responses to cardiac transplantation and coronary artery bypass graft surgery. J Thorac Cardiovasc Surg 1996;111:469-77.
- 124. Shibatta M, Endo S, Inada K, Kuriki S, Harada M, Takino T, et al. Elevated plasma levels of interleukin-1 receptor antagonist and interleukin-10 in patients with acute myocardial infarction. J Interferon Cytokine Res 1997;92:1-5.
- 125. Yang Z, Zingarelli B, Szabo C. Crucial role of endogenous interleukin-10 production in myocardial ischaemia/reperfusion injury. Circulation 2000;101:1019-26.
- 126. Zhai QH, Futrell N, Chen FJ. Gene expression of IL-10 in relationship to TNF-alpha, IL-1beta, and IL-2 in the rat brain following middle cerebral artery occlusion. J Neurol Sci 1997;152:

Pérez Fernández R, et al. Interleukin-10 and Coronary Disease

119-24.

- 127. Le Moine O, Louis H, Stordeur P, Collet JM, Goldman M, Deviere J. Role of reactive oxygen intermediates in interleukin 10 release after cold ischaemia liver ischaemia and reperfusion in mice. Gastroenterology 1997;113:1701-6.
- 128. Hayward R, Nossuli TO, Scalia R, Lefer AM. Cardioprotective effect of interleukin-10 in murine myocardial ischaemia-reperfusion. Eur J Pharmacol 1997;334:157-63.
- 129. Hess PJ, Seeger JM, Huber TS, Welborn MB, Martin TD, Harward TR, et al. Exogenously administrated interleukin-10 decreases pulmonary neutrophil infiltration in a tumor necrosis factor-dependent murine model of acute visceral ischaemia. J Vasc Surg 1997;26:113-18.
- 130. Engles RE, Huber TS, Zander DS, Hess PJ, Welborn MB, Moldawer LL, et al. Exogenous recombinant interleukin-10 attenuates hindlimb ischaemia-reperfusion injury. J Surg Res 1997;69: 425-8.
- 131. Spera PA, Ellison JA, Feuerstein GZ, Barone FC. IL-10 reduces rat brain injury following focal stroke. Neurosci Lett 1998;251:189-92.
- 132. Huang S, Ullrich SE, Bar-Eli M. Regulation of tumor growth and metastasis by interleukin-10: the melanoma experience. J Interferon Cytokine Res 1999;19:697-703.
- 133. Stearns ME, Garcia FU, Fudge K, Rhim J, Wang M. Role of interleukin 10 and transforming growth factor $β_1$ in the angiogenesis and metastasis of human prostate primary tumor lines from orthotopic implants in severe combined immunodeficiency mice. Clin Cancer Res 1999;5:711-20.
- 134. Carmeliet P. Mechanisms of angiogenesis and arteriogenesis. Nat Med 2000;6:389-95.
- 135. Silvestre JS, Mallat Z, Duriez M, Tamarat R, Bureau MF, Scherman D, et al. Antiangiogenic effect of interleukin-10 in is-

chaemia-induced angiogenesis in mice hindlimb. Cir Res 2000; 87:448-52.

- 136. Terkeltaub RA. Il-10: an «immunologic scalpel» for atherosclerosis? Arterioscler Thromb Vasc Biol 1999;19:2823-5.
- 137. Rogy MA, Beinhauer Bg, Reinisch W, Huang L, Pokieser P. Transfer of interleukin-4 and interleukin-10 in patients with severe inflammatory bowel disease of the rectum. Hum Gene Ther 2000;11:1731-41.
- 138. Fedorak RN, Gangl A, Elson CO, Rutgeerts P, Schreiber S, Wild G, et al. Recombinant Human Interleukin 10 in the treatment of patients with mild to moderately active Crohn's disease. Gastroenterology 2000;119:1473-82.
- 139. Colombel JF, Rutgeerts P, Malchow H, Jacyna M, Nielsen OH, Rask-Madsen J, et al. Interleukin 10 (Tenovil) in the prevention of postoperative recurrence of Crohn's disease. Gut 2001; 49:42-6.
- 140. Bickston S, Cominelli F. Recombinant interleukin 10 for the treatment of active Crohn disease: lessons in biologic therapy. Gastroenterology 2000;119:1781-3.
- 141. Kühn R, Löhler J, Rennick D, Rajewsky K, Muller W. Interleukin-10-deficient mice develop chronic enterocolitis. Cell 1993;75:263-74.
- 142. Hagenbaugh A, Sharma S, Dubinett SM, Wei SH, Aranda R, Cheroutre H, et al. Altered immune responses in interleukin 10 transgenic mice. J Exp Med 1997;185:2101-10.
- 143. Yang X, HayGlass KT, Brunham RC. Genetically determined differences in IL-10 and IFN-gamma responses correlate with clearance of *Chlamydia trachomatis* mouse pneumonitis infection. J Immunol 1996;156:4338-44.
- 144. Yang X, Gartner J, Zhu L, Wang S, Brunham RC. IL-10 gene knockout mice show enhanced Th1-like protective immunity and absent granuloma formation following Chlamydia trachomatis lung infection. J Immunol 1999;162:1010-7.